

gamma and PKC delta dependent and proceeds via a mitochondrial pathway.

Disclosures:

The following people have nothing to disclose: Andrea N. Johnston, Kathleen Ponzetti, Simon Hohenester, Mohammed S. Anwer, Cynthia R. Webster

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CARBON TETRACHLORIDE-INDUCED LIVER INJURY AND FIBROSIS CORRELATES WITH OSTEOPOINTIN EXPRESSION IN MICE

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Background: Osteopontin (OPN) is a multifunctional matricellular cytokine that plays a significant role in innate immunity, cell survival, tumor invasion, and metastasis. **Aim:** We have previously shown that OPN promotes activation of quiescent hepatic stellate cells and increases collagen I expression and secretion. Here, we elucidated the role of OPN in the pathogenesis of hepatic fibrosis *in vivo* using both OPN transgenic mice (*Opn* Tg) and OPN knockout mice (*Opn*^{-/-}). **Methods:** Liver fibrosis was induced in C57BL/6 WT, *Opn* Tg and *Opn*^{-/-} mice by i.p. injections of carbon tetrachloride twice a week for 1 month (5 μ l CCl₄/10 g b. wt.), which induces significant oxidative stress via generation of CCl₃ radical. Commercially available kits were used for biochemical assays. H&E staining and immunohistochemistry were carried out to determine the extent of liver injury. Samples were scored by an experienced hepatopathologist. **Results:** To decipher the role of OPN in progressive liver injury, we tested whether liver injury and fibrosis under chronic CCl₄ administration could correlate with OPN expression. WT mice under CCl₄ treatment showed marked elevation of serum AST, ALT and γ GT, along with striking hepatic inflammation, necrosis, ballooning, activity score, activation of hepatic stellate cells, and scarring. All these pathophysiological markers were significantly elevated by CCl₄ in *Opn* Tg mice but were attenuated in *Opn*^{-/-} mice compared to WT mice. There was up-regulation of collagen I and OPN proteins in CCl₄-treated *Opn* Tg mice, while the opposite occurred in *Opn*^{-/-} mice, compared to CCl₄-treated WT mice. *Opn* Tg mice injected with CCl₄ showed elevated collagenous proteins, portal fibrosis, bridging fibrosis, greater collagen I band thickness, and fibrosis score than CCl₄-injected WT or *Opn*^{-/-} mice. Immunohistochemical analysis revealed massive induction of OPN in biliary epithelial cells, oval cells, and hepatic stellate cells under CCl₄ treatment in WT and *Opn* Tg mice. OPN+ cells were organized in small nests or arborizing duct-like structures, while isolated cells were found at some distance from portal tracts. **Conclusion:** These results suggest that OPN plays a significant role in the pathogenesis of hepatic fibrosis *in vivo*; thus, opening up the possibility of blocking OPN for preventing the development of liver fibrosis.

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DELETION OF C-FLIP IN HEPATOCYTES AUGMENTS CCL4-INDUCED LIVER INJURY AND FIBROSIS

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The mechanisms that contribute to chronic liver injury and promote liver fibrosis are only incompletely understood. The caspase-8 homologue cellular FLICE inhibitory protein (cFLIP) acts to protect cells from apoptosis involving impaired activation of pro-caspase 8 at the level of the death inducing signaling complex (DISC). We have previously shown that deletion of c-FLIP enhances hepatocellular apoptosis from activation of receptors of the TNF-receptor superfamily. Based on these observations, we hypothesized that deletion of c-FLIP would augment acute and chronic liver injury and fibrosis from the hepatotoxin CCl₄. To test this hypothesis mice exhibiting a hepatocyte-specific deletion of c-FLIP were generated using the cre-loxP system under control of an albumin promoter (Δ hepFLIP). To induce liver injury wild type (wt) and Δ hepFLIP mice were treated with CCl₄ for a total of 8 weeks and liver injury was assessed at an early (acute injury) and late (chronic injury) stage through measurement of serum ALT. We observed increased ALT in Δ hepFLIP at both time points. At 24h Δ hepFLIP mice exhibited 2,5-fold higher serum transaminases compared to wt mice (ALT: 2865 \pm 425 vs. 7540 \pm 2804 wt vs. Δ hepFLIP, p=0,05). Following 8 weeks of treatment, the absolute levels of ALT were slightly lower, however liver injury from CCl₄ was still significantly higher in Δ hepFLIP mice compared to wt mice (ALT: 3317 \pm 690 vs. 4914 \pm 709, wt vs. Δ hepFLIP). Histological analysis of H&E and Goldner-stained liver sections revealed that Δ hepFLIP mice exhibited significantly more necrotic hepatocytes and hepatic fibrosis. Additionally, transcription of collagen I, measured by real time-PCR was increased by 2.6 compared to wt mice. Since interleukin 6 (IL6) was previously shown to augment liver injury and fibrosis from CCl₄ we examined mRNA levels of this pro-inflammatory cytokine and found that IL6 was increased 6-fold in Δ hepFLIP mice compared to wt mice. In summary, deletion of c-FLIP in hepatocytes augments CCl₄-induced acute and chronic liver injury and hepatic fibrosis potentially involving increased levels of IL6. These findings imply that c-FLIP could potentially contribute to liver regeneration and fibrosis in addition to the regulation of cell death pathways.

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